UNIVERSITÀ DI PISA

Centro E. Piaggio
bioengineering and robotics research center

INGEGNERIA DEI TESSUTI BIOLOGICI:

DYNAMIC MECHANICAL ANALYSIS (DMA)

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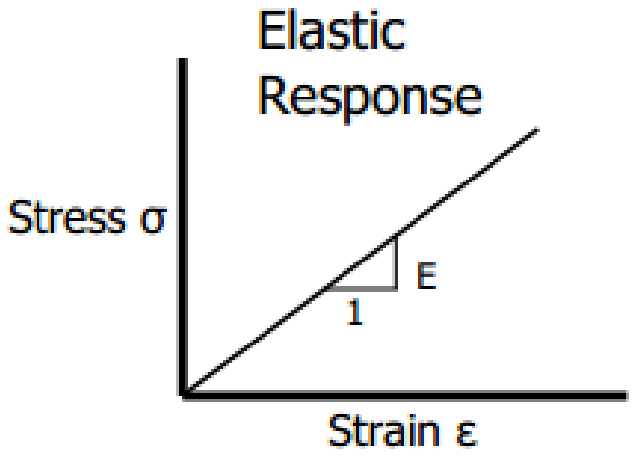
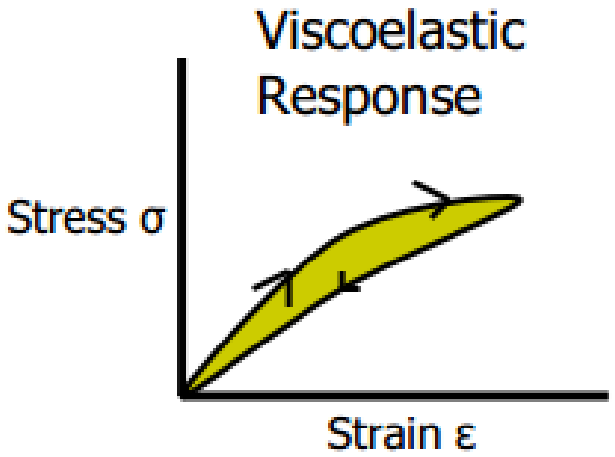
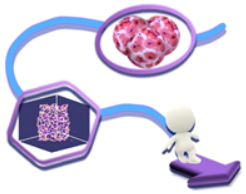
Viscoelasticity

- **Viscoelastic materials** exhibit the characteristics of both elastic and viscous materials
  - Viscosity → resistance to flow (damping)
  - Elasticity → ability to revert back to the original shape
- **Elastic vs. viscoelastic** stress-strain response







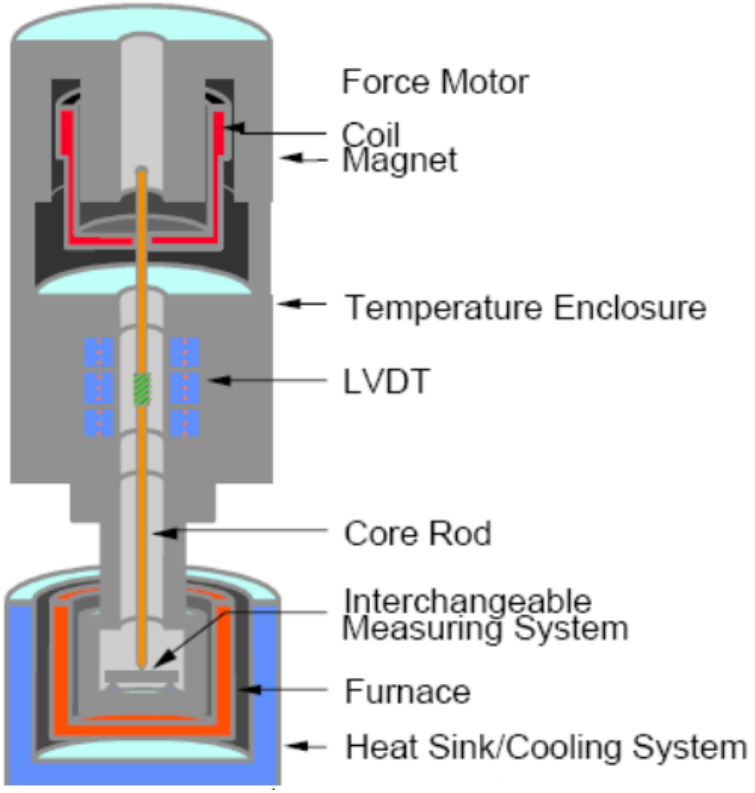
Methods to characterise viscoelasticity

- **Time** domain
  - Creep response
  - Stress relaxation
  - Epsilon dot Method ($\dot{\varepsilon}M$, Tirella A. et al., JBMR 2013)

- **Frequency** domain
  - Dynamic mechanical analysis (DMA)
  - Dynamic mechanical thermal analysis (DMTA)

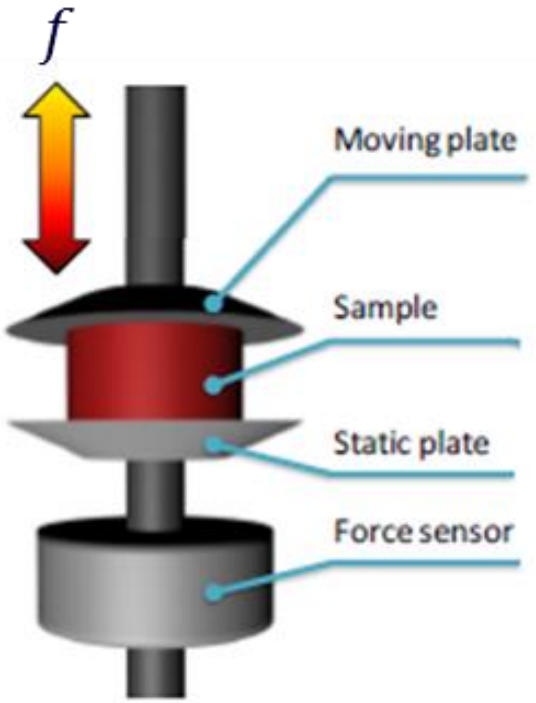
DMA overview

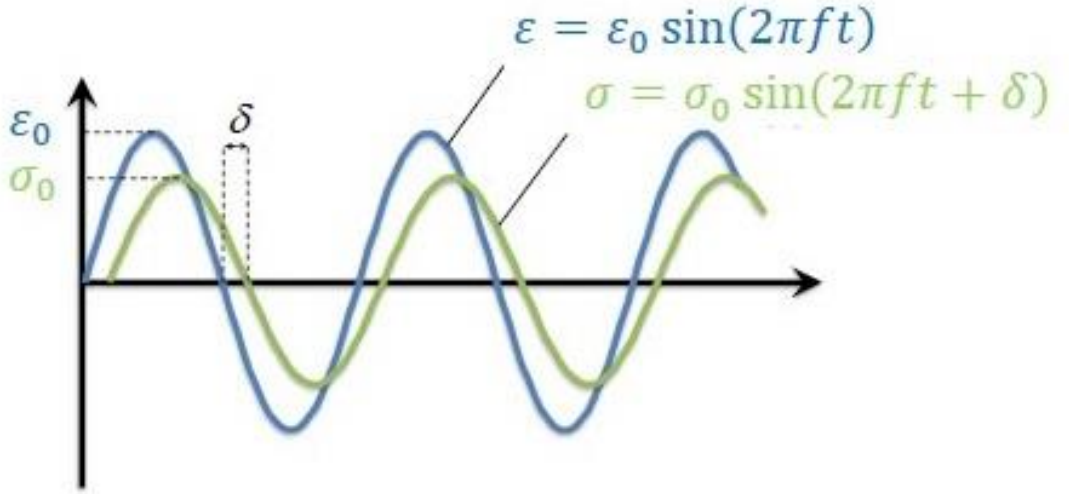
- Dynamic mechanical analysis (DMA) is a standard **force-triggered method** to **determine viscoelastic properties** of materials by **applying a small amplitude cyclic strain** on a sample and **measuring the resultant cyclic stress response**.



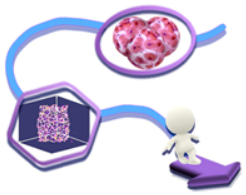
DMA overview

- For a given **sinusoidal strain input** the resulting **stress will be sinusoidal if the applied strain is small enough** so that the tissue can be approximated as linearly viscoelastic.



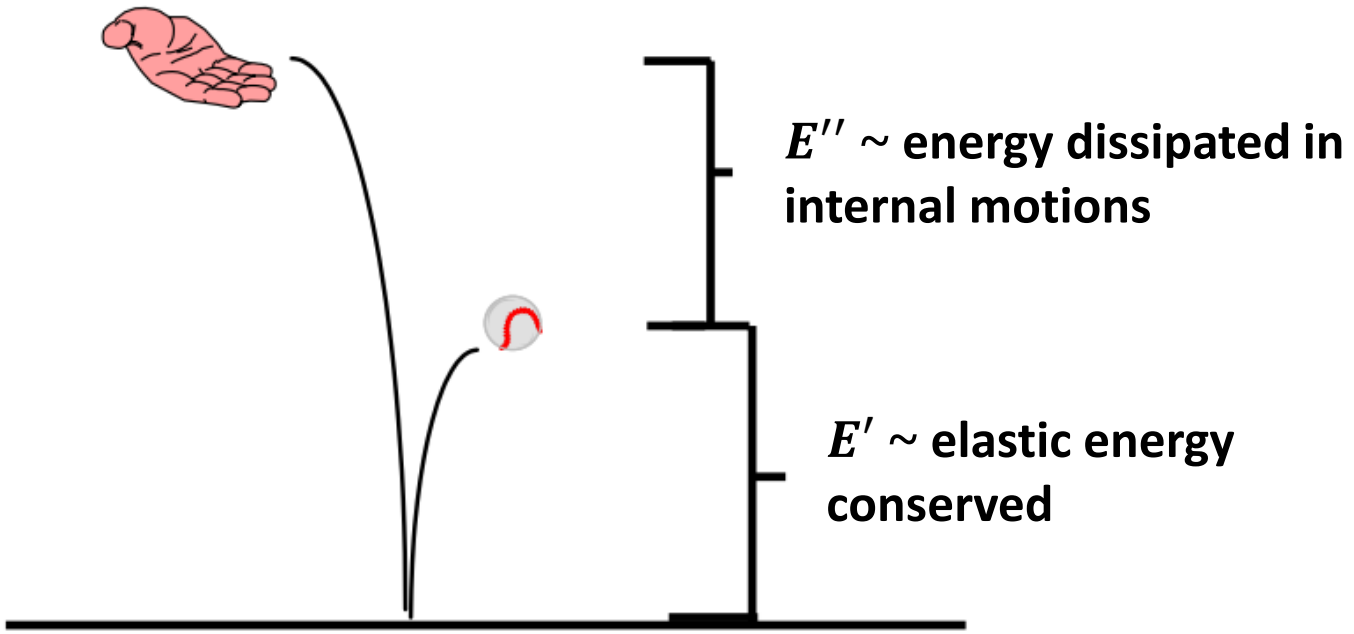


Viscoelastic material response is characterised by a **phase lag** (**δ**) between the strain input and the stress response, which is comprised **between 0°** (**purely elastic**) and **90°** (**purely viscous**). This phase lag is **due to** the excess **time necessary for molecular motions and relaxations** to occur.



Complex, storage and loss modulus

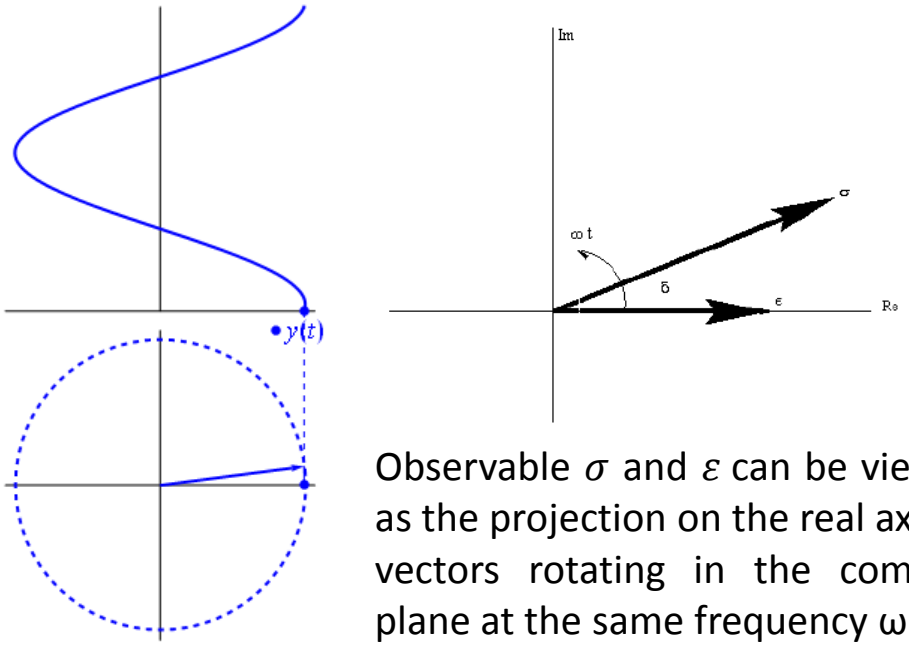
- The dynamic mechanical properties are quantified with the **complex modulus** ($\boldsymbol{E^*}$), which can be thought as an **overall resistance** to deformation under dynamic loading. The complex modulus is composed of the **storage** ($\boldsymbol{E'}$, elastic component) and the **loss** ($\boldsymbol{E''}$, viscous component) moduli, that are **additive under the linear theory of viscoelasticity** ($\boldsymbol{E^* = E' + iE''}$).



Definitions

- It is convenient to represent the sinusoidal stress and strain functions as complex quantities (called rotating vectors, or **phasors**) with a **phase shift** of **δ**.

$$\varepsilon = \varepsilon_0 \, e^{i\omega t} \qquad \sigma = \sigma_0 \, e^{i(\omega t+\delta)}$$



Observable σ and ε can be viewed as the projection on the real axis of vectors rotating in the complex plane at the same frequency ω

Rotating vector representation of harmonic stress and strain

$$\boldsymbol{E}^* = \frac{\sigma}{\varepsilon} = \frac{\sigma_0}{\varepsilon_0} e^{i\delta} =$$

$$= \frac{\sigma_0}{\varepsilon_0} (\cos\delta + i \sin\delta) =$$

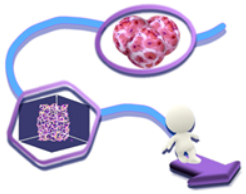
$$= \boldsymbol{E'} + \boldsymbol{iE''}$$

Storage modulus $E' = E^* \cos(\delta)$

Loss modulus $E'' = E^* \sin(\delta)$

$\tan(\delta) = E''/E'$ Damping factor

$\eta' = E''/\omega$ Dynamic viscosity



Test modes

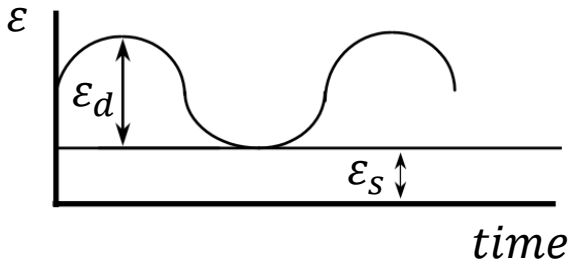
- **Temperature sweep**: Modulus and damping are recorded as the sample is heated
- **Frequency sweep**: Modulus and damping are recorded as the sample is loaded at increasing (or decreasing) frequencies
- **Stress amplitude sweep**: Modulus and damping are recorded as the sample stress is increased
- **Strain amplitude sweep**: Modulus and damping are recorded as the sample strain is increased
- **Combined sweep**: Combinations of above methods

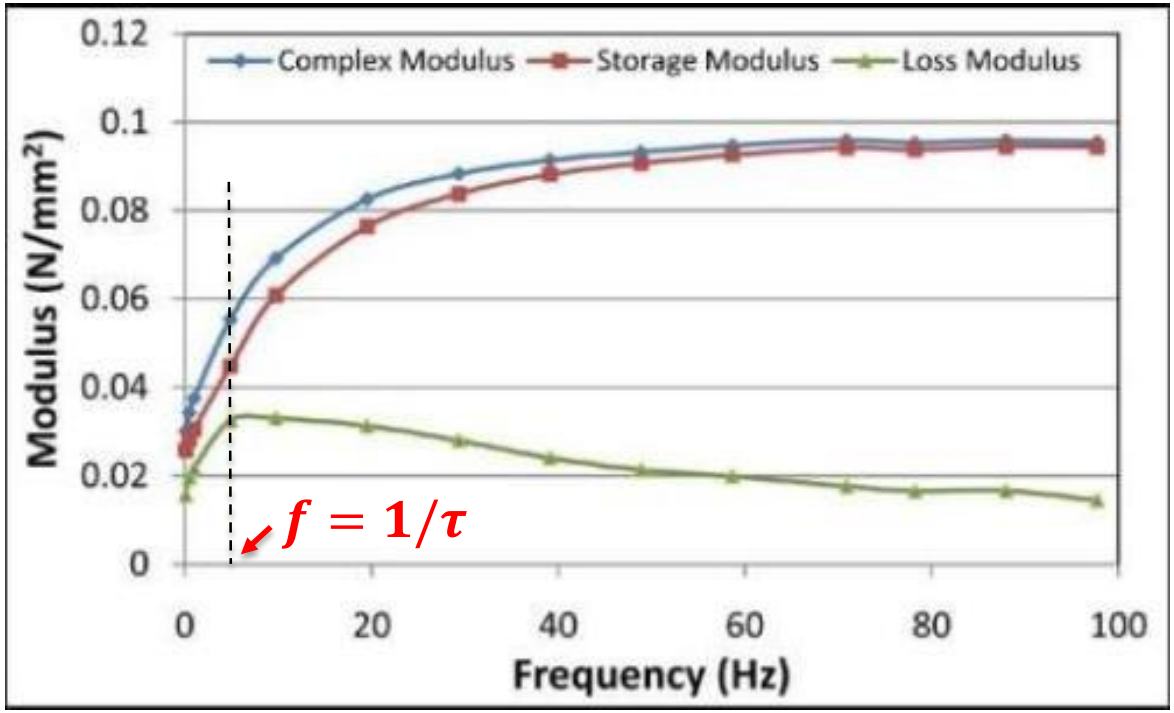
Frequency sweep tests

- A sample is held to a **fixed temperature** and tested at **varying frequency**.

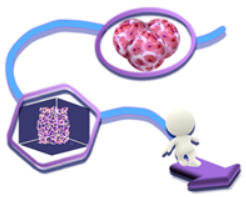
Test parameters:

- Temperature (T)
- Frequency range (*f*)
- Static strain (ε_s)
- Dynamic strain (ε_d)



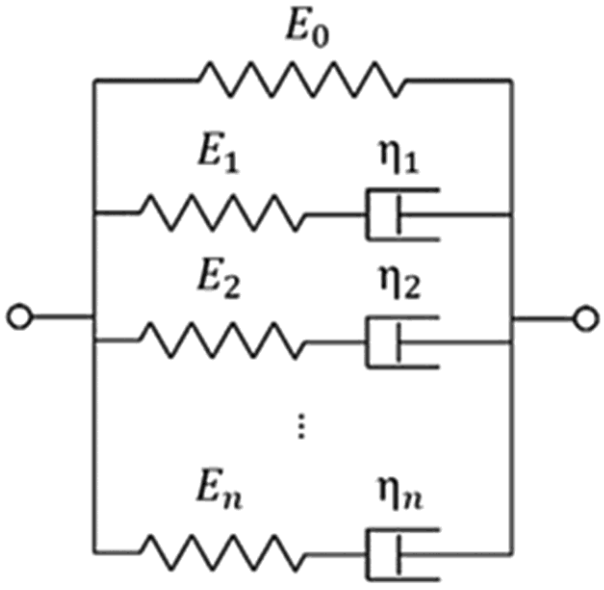


- **Peaks** in $\tan(\delta)$ or E'' with respect to frequency identify the **characteristic relaxation frequencies** of the viscoelastic sample under testing, defined as $f = 1/\tau$, where τ is the **characteristic relaxation time)**



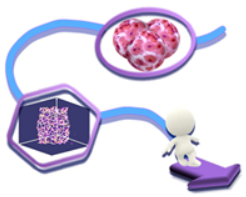
Lumped models to describe material linear viscoelastic response

- The most general form of linear viscoelastic model is called the **Generalised Maxwell (GM)** model and consists of a **pure spring (E_0)** with n **Maxwell arms** (i.e. spring E_i in series with a dashpot η_i) assembled **in parallel**, thus defining a set of n **different characteristic relaxation times** (i.e. $\tau_i = \eta_i / E_i$)



$$H_{GM}(s) = \frac{\bar{\sigma}}{\bar{\epsilon}} = E_0 + \sum_{i=i}^{n} \frac{E_i \, \eta_i \, s}{E_i + \eta_i \, s}$$

GM model transfer function in the Laplace domain

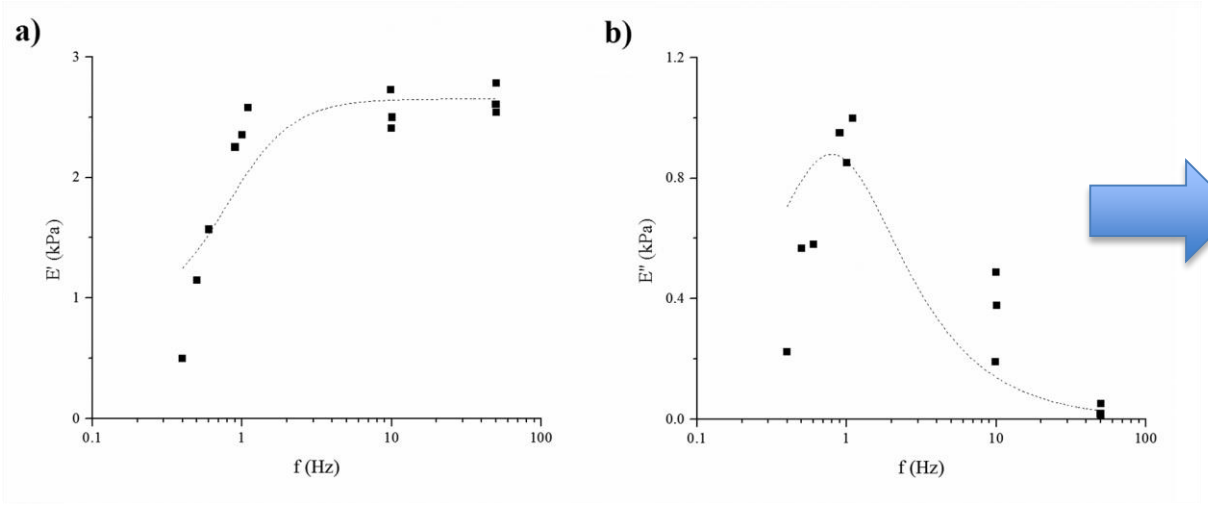


Lumped parameters derivation from frequency sweep tests

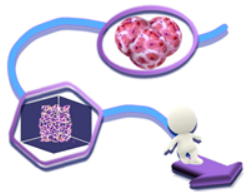
- Calculate the **complex conjugate of the GM modulus** (E^*_{GM}) by substituting $s = i\,\omega = i\,2\pi f$ in $H_{GM}(s)$, then **split the expression into** its **real (*Re*)** and **imaginary (*Im*)** parts to obtain the **frequency-dependent relations** for the **storage** and **loss** moduli, respectively

$$E^*_{GM}(f) = \underbrace{\left(E_0 + \sum_{i=i}^{n} \frac{4\,E_i\,\eta_i^2\,f^2\,\pi^2}{E_i^2\ +\ 4\,\eta_i^2 f^2 \pi^2}\right)}_{E'(f)} + i\underbrace{\left(\sum_{i=i}^{n} \frac{2\,E_i^2\,\eta_i\,f\,\pi}{E_i^2\ +\ 4\,\eta_i^2 f^2 \pi^2}\right)}_{E''(f)}$$

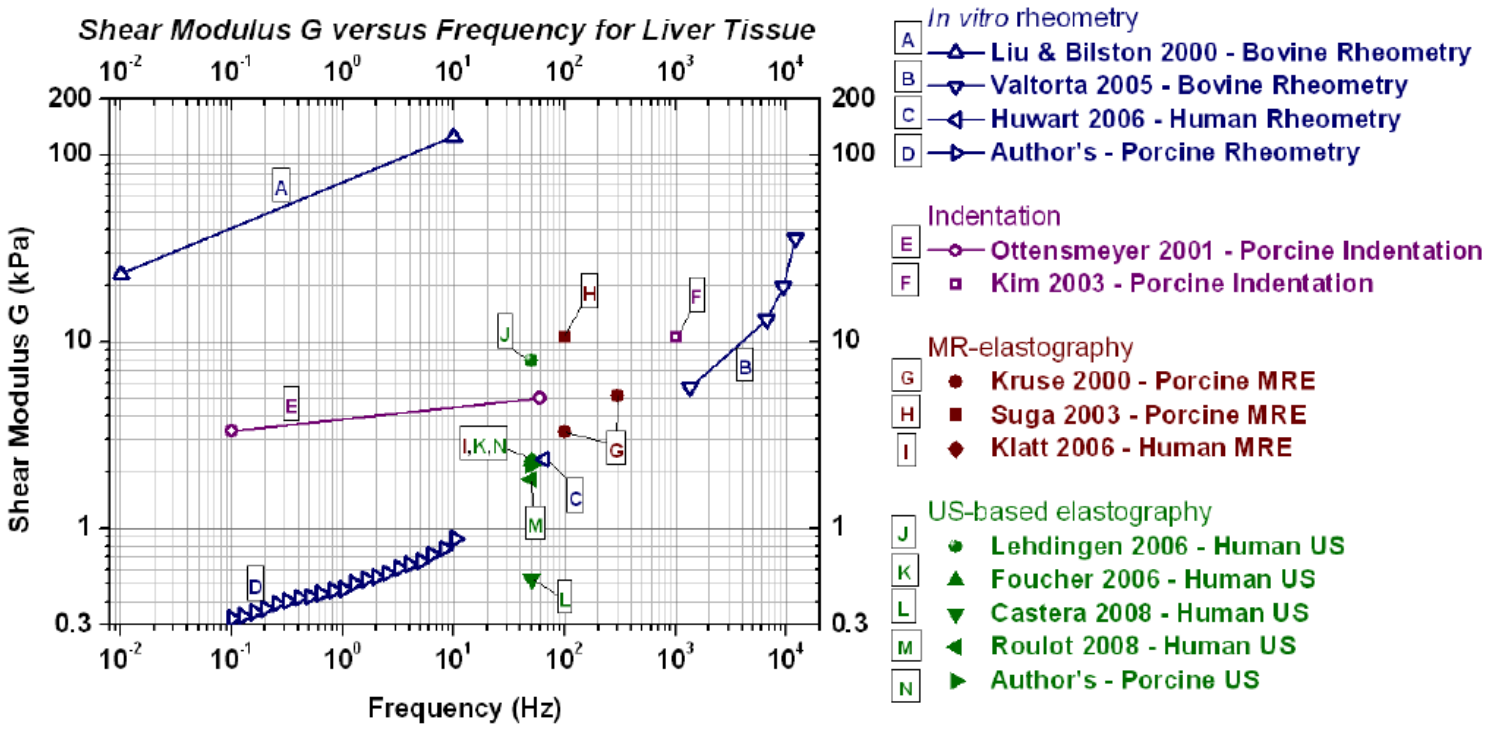
- **Global fitting** with **shared parameters** (χ^2 minimisation)



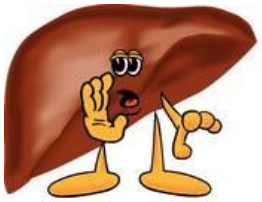
CASE OF STUDY:
THE LIVER



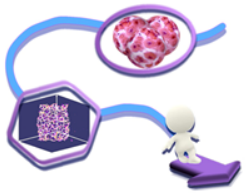
SoA: a myriad of different results



Source: S. Marchesseau et al., *Progress in biophysics and molecular biology*, 103:2–3, pp. 185–96, 2010



Many **variables and factors affect** measured **liver mechanical properties,** leading to a **lack of consensus and unique properties**, which are **critical for developing appropriate viscoelastic models**

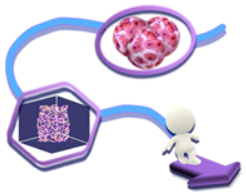


Typical variability factors

- **Testing condition**
  - ***in-vivo****: tissue in its* ***natural state****, but many* ***testing limitations***
  - ***ex-vivo:*** *better for* ***developing testing devices****,* ***protocols*** *and* ***tissue models***
- **Testing method and experimental setup**
  - **Direct measurements** or **image-based techniques**
  - **Time, strain rate** or **frequency range** considered
- **Tissue sample**
  - **Type and source:** animal source, presence of Glisson's capsule
  - **Status:** environmental testing parameters, physical conditions, post-mortem time, preservation period, pathophysiological state, preload

From this multifaceted research area emerges **the need to:**

1. **clearly identify the parameters of interest**
2. **develop suitable experimental testing setup and protocols** for the **unique identification** of **liver viscoelastic parameters**

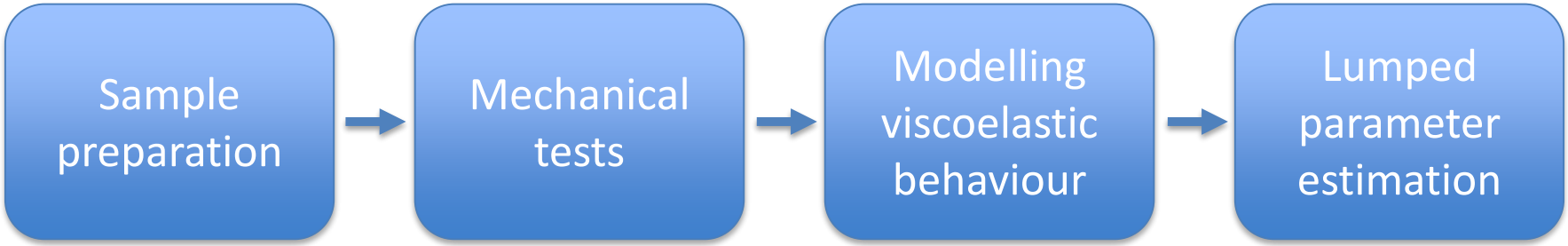


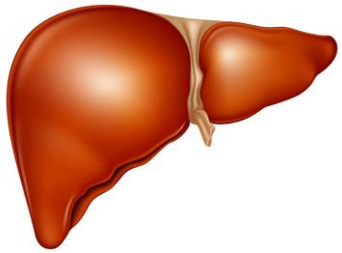
Aim and strategy

AIM: establishing an **experimental testing and analysis framework** to **unequivocally characterise** the **liver viscoelastic behaviour** in the **LVR** (linear viscoelastic region)

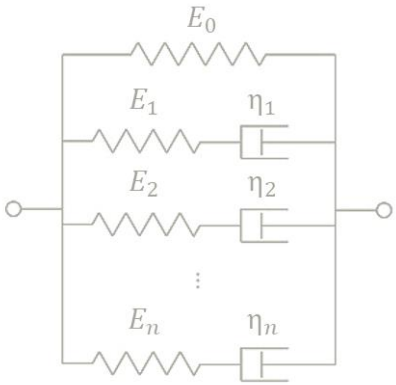
STRATEGY: ***ex-vivo*** measurements in **unconfined compression** using **common testing apparatus** and **2 different testing methods**

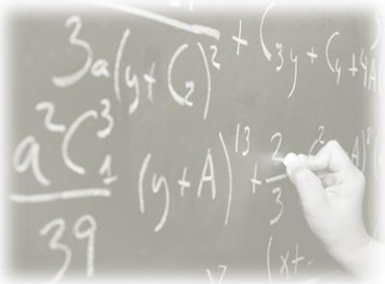
- $\dot{\varepsilon}M$, **a solution** to **avoid major drawbacks** of force- or strain-triggered methods **in testing floppy samples** (e.g. **long test duration** and significant **sample pre-load**)
- ***step-reconstructed* DMA**, a modification of a **widely used technique** for viscoelastic characterisation of materials





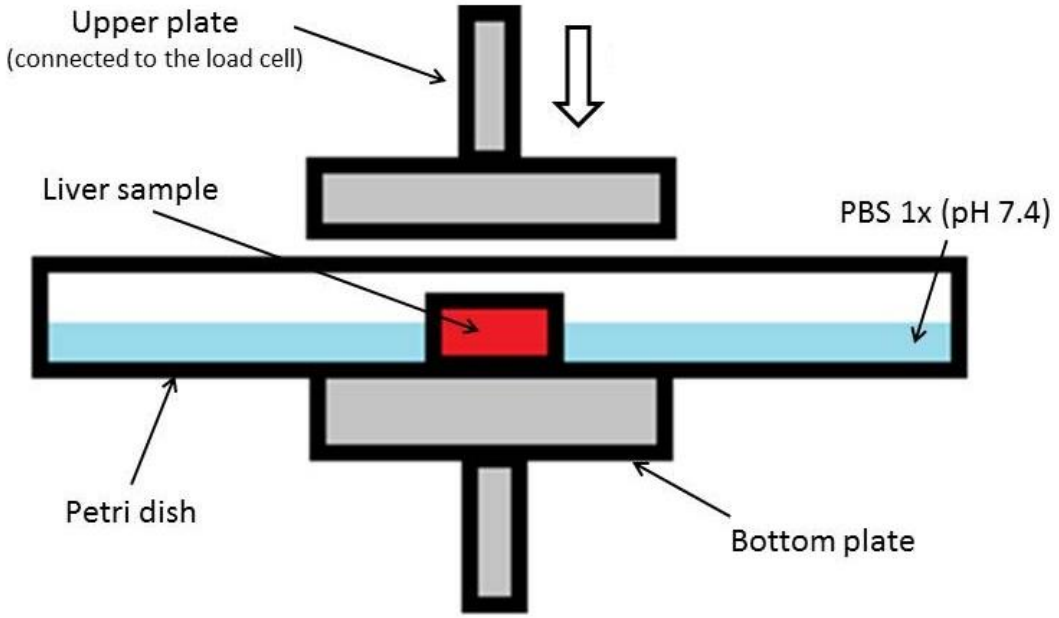
Sample preparation



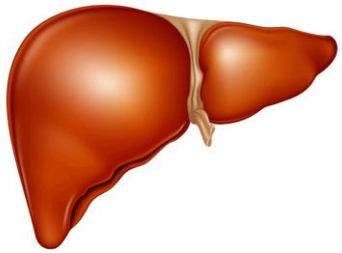


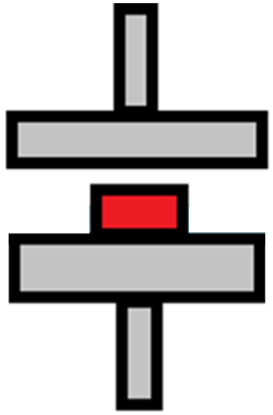
Sample preparation and testing configuration

- **Cylindrical liver samples** (14 mm diameter, 3 mm thickness) collected from **1 year old healthy pigs avoiding Glisson's capsule** and **macroscopic vasculature**
- **Repeatable testing condition** → samples **equilibrium swollen in PBS 1x at 4°C**, then **brought to room T** and **carefully measured** prior testing

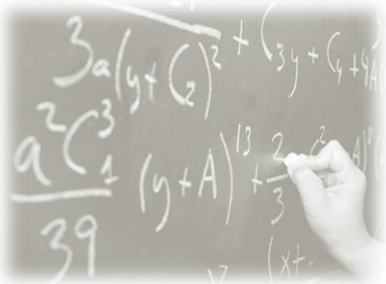


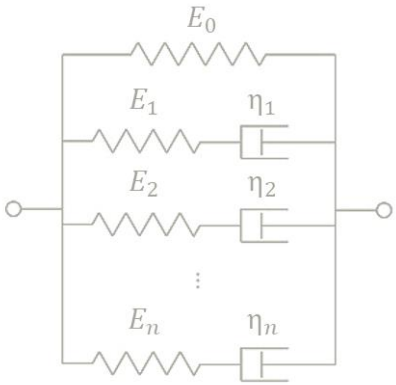
Testing configuration





Mechanical tests

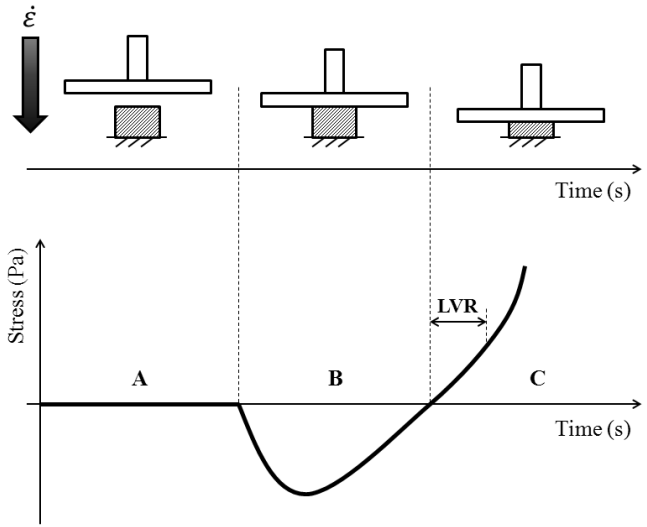




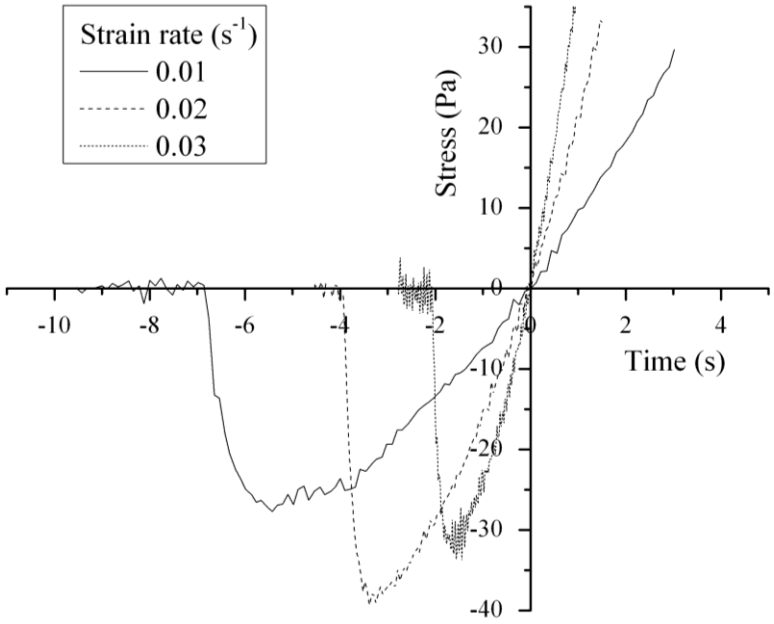
$\dot{\varepsilon}M$: short test with no pre-load

A. Tirella, G. Mattei, A. Ahluwalia, JBMR Part A (2013)

$\dot{\varepsilon}M$ paradigm: characterise the material viscoelastic behaviour testing samples at **different constant strain rates** ($\dot{\varepsilon}$), then **analysing $\sigma(t)$ curves**



- ✓ Implementable with **all uniaxial testing devices**
- ✓ **Force-displacement time recording** starts **prior to** sample **contact** → **no pre-load**
- ✓ **Short test** duration → **no sample deterioration**
- ✓ **LVR** determined through **measured σ-ε curves**
- x Need **preliminary tests** or an ***a priori* knowledge of the material relaxation behaviour** to **choose $\dot{\varepsilon}$**



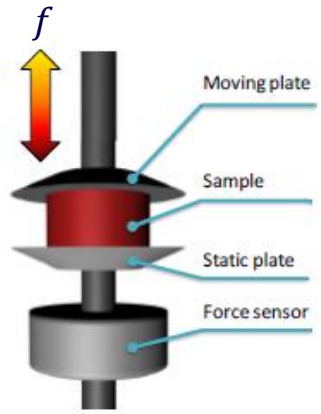
Experimental stress-time data at various $\dot{\varepsilon}$
(only **LVR values** are shown in zone C)

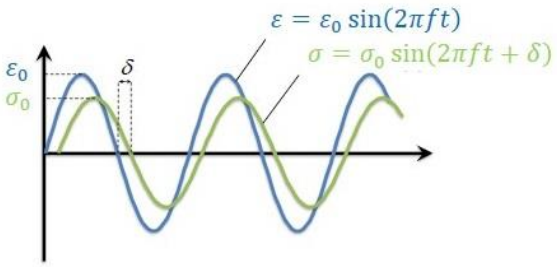
Zwick/Roell Z005, 10N load cell

3 samples x *3 $\dot{\varepsilon}$ = 9 samples*

DMA: a widely accepted method

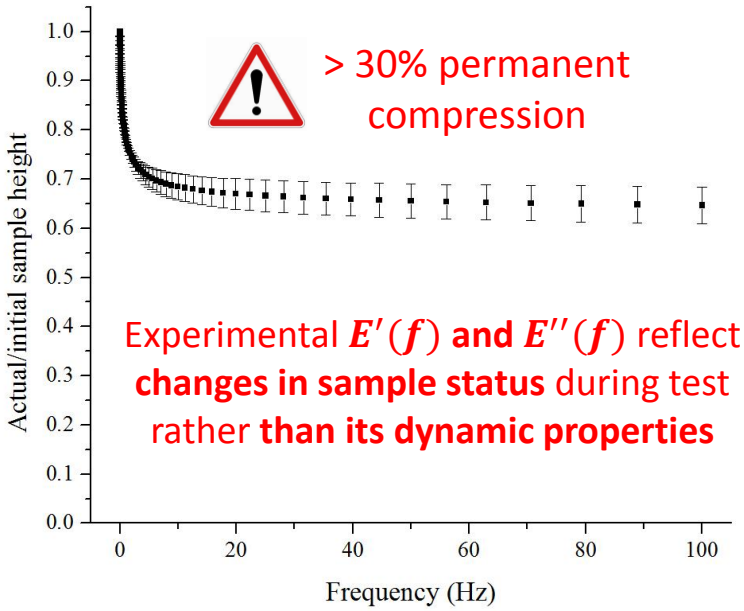
DMA paradigm: characterise viscoelastic behaviour testing samples at **different frequencies** (f), then **analysing** $E'(f)$ **and** $E''(f)$





$$E' = \frac{\sigma_0}{\varepsilon_0}\cos(\delta) \qquad E'' = \frac{\sigma_0}{\varepsilon_0}\sin(\delta) \qquad E^* = E' + iE''$$

- ✓ **Largely accepted** for **viscoelastic characterisation**
- ✓ Wide **frequency sweep** tests **simplify testing set-up** avoiding preliminary tests or any *a priori* knowledge
- x **Long testing time** may **degrade the sample**
- x **Trigger force** may **significantly pre-load samples**
- x **Preliminary strain-sweep tests** to **derive the LVR**

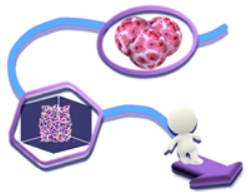


Experimental $E'(f)$ **and** $E''(f)$ reflect **changes in sample status** during test rather **than its dynamic properties**

Permanent deformation during a 0.05 – 100 Hz frequency sweep test (~ **1.5 h**)

GABO Eplexor 150N, ***10mN*** *trigger force*

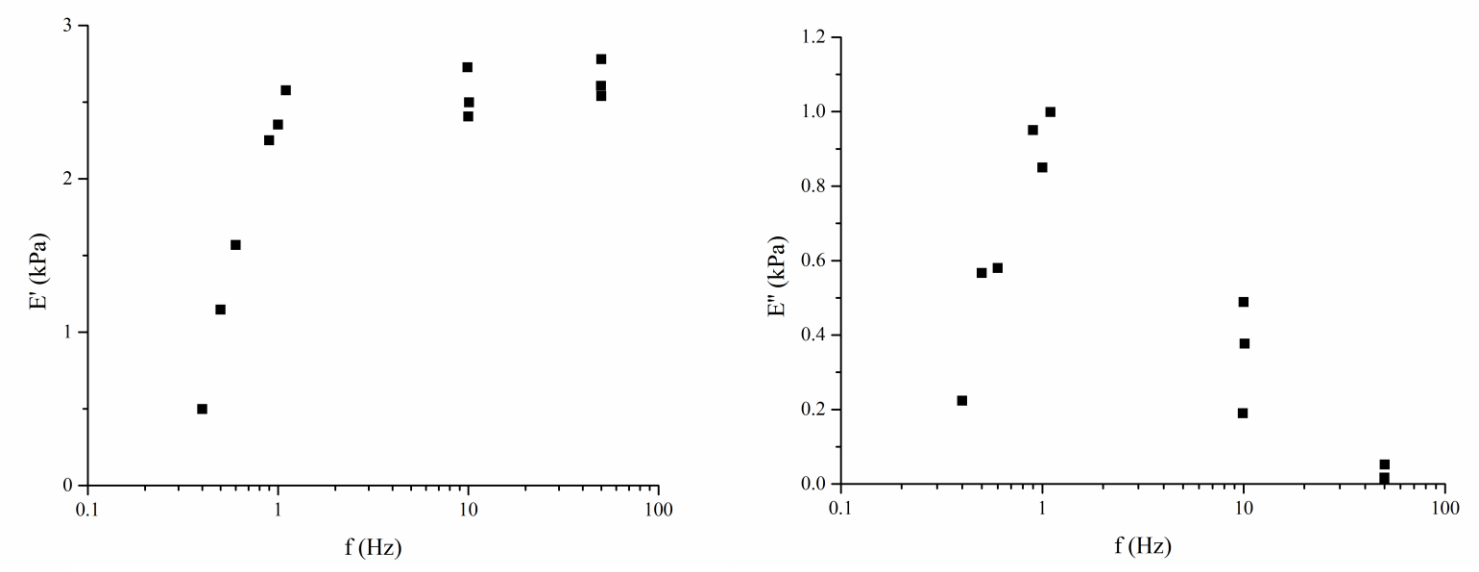
3 samples



step-reconstructed (SRDMA)

G. Mattei, A. Tirella, G. Gallone, A. Ahluwalia, *submitted*

SRDMA paradigm: perform **DMA measurements around specific *f*,** then **reconstruct $E'(f)$ and $E''(f)$ over** the whole **frequency range of interest**



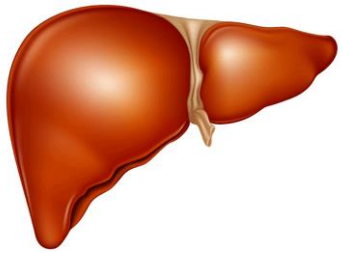
Storage (E') and loss (E'') moduli measured around *f* = 0.5, 1, 10 and 50 Hz (*f* - 0.1 Hz, *f*, *f* + 0.1 Hz)

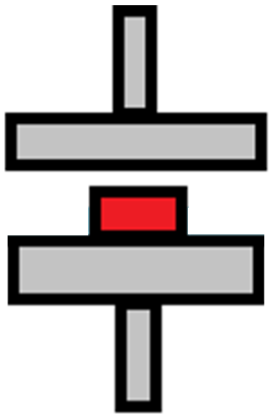
*GABO Eplexor 150N, **10mN** trigger force*
3 samples x *4 f = 12 samples*

✓ **Short testing time** → **no sample deterioration** (**< 2 %** permanent compression in the *worst* case, i.e. *f* = 0.5 Hz)

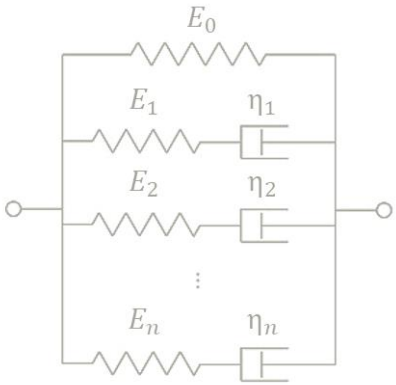
x **Trigger force** → **sample pre-load**

x Need **preliminary tests** or an ***a priori* knowledge of the material relaxation behaviour** to **choose *f***

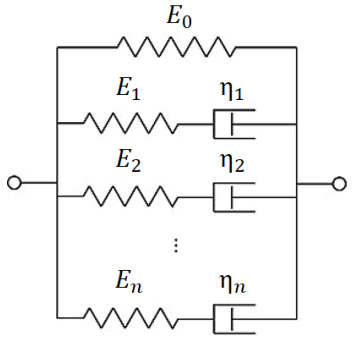




Modelling viscoelastic behaviour



Generalised Maxwell (GM) model

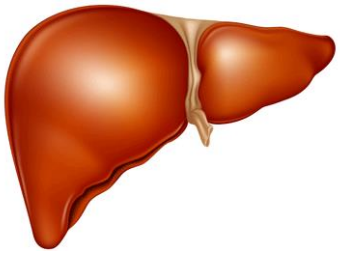


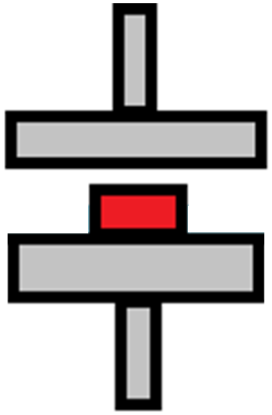
$\tau_i = \eta_i / E_i$ i^{th} relaxation time

$$H_{GM}(s) = \frac{\bar{\sigma}}{\bar{\epsilon}} = E_0 + \sum_{i=i}^{n} \frac{E_i \, \eta_i \, s}{E_i + \eta_i \, s}$$

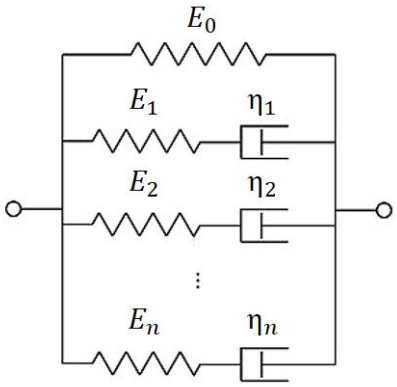
Transfer function in the **Laplace** domain

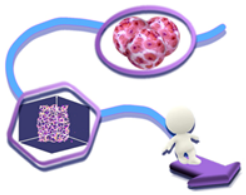
	$\dot{\varepsilon}M$ needs $\sigma(t)$ response to a fixed $\dot{\varepsilon}$	SRDMA needs $E'(f)$ and $E''(f)$
General form	$\bar{\sigma} = H_{GM}(s) \cdot \frac{\lvert\dot{\varepsilon}\rvert}{s^2}$ ← **Laplace transform** of a **constant $\dot{\varepsilon}$ input** with **amplitude** $\lvert\dot{\varepsilon}\rvert$ **Inverse Laplace transformation**	$E^*_{GM}(f) = \underbrace{\left(E_0 + \sum_{i=i}^{n} \frac{4 E_i \eta_i^2 f^2 \pi^2}{E_i^2 + 4 \eta_i^2 f^2 \pi^2}\right)}_{E'(f)} + i \underbrace{\left(\sum_{i=i}^{n} \frac{2 E_i^2 \eta_i f \pi}{E_i^2 + 4 \eta_i^2 f^2 \pi^2}\right)}_{E''(f)}$
Max SLS ($n = 1$)	$\sigma(t) = \dot{\varepsilon}\left[E_0 t + \eta_1 (1 - e^{-\frac{E_1}{\eta_1} t})\right]$	substitute $n = 1$ in the general equation
GM2 ($n = 2$)	$\sigma(t) = \dot{\varepsilon}\left[E_0 t + \eta_1 (1 - e^{-\frac{E_1}{\eta_1} t}) + \eta_2 (1 - e^{-\frac{E_2}{\eta_2} t})\right]$	substitute $n = 2$ in the general equation





Lumped parameter estimation



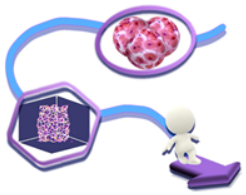


Global fitting with shared parameters

$\dot{\varepsilon}M$	SRDMA
1. Choose a **lumped parameter model**	
2. Calculate $\sigma(t)$ response to a **fixed $\dot{\varepsilon}$**	**2. Calculate $E'(f)$** and **$E''(f)$**
3. Build a **unique dataset** for the **global fit** and **share** the **viscoelastic parameters**	
4. Fix $\dot{\varepsilon}$ in the fitting equation of **each experimental $\sigma(t)$** to the **applied $\dot{\varepsilon}$**	**4. Associate exp. data** to the **modelled expressions** of **$E'(f)$** and **$E''(f)$**
5. Global fit performing **χ^2 minimisation** in a **combined parameter space**	

Annealing scheme
to avoid most of the local minima

Viscoelastic constants (E_i, η_i) for the **chosen model**



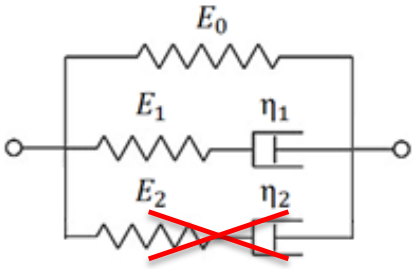
Global fitting results

Porcine liver viscoelastic parameters (estimated value ± standard error)

	Maxwell SLS		GM2	
Parameter	$\dot{\varepsilon}M$	**SRDMA**	$\dot{\varepsilon}M$	**SRDMA**
E_{inst} (kPa)	2.04 ± 0.01	2.04 ± (3.21 · 10^2) *n.s.*	2.65 ± 0.30	2.65 ± (3.61 · 10^5) *n.s*
E_{eq} (kPa)	0.91 ± 0.01	0.91 ± 0.01	0.89 ± 0.22	0.89 ± 0.56
τ_1 (s)	1.10 ± 0.02	1.10 ± (3.05 · 10^3) *n.s.*	0.20 ± 0.06	0.20 ± (1.14 · 10^5) *n.s.*
τ_2 (s)	-	1.10 ± (3.43 · 10^2) *n.s.*	-	0.20 ± (0.65 · 10^5) *n.s.*
R^2	0.97	0.97	0.92	0.92

n.s. → *non significant estimate*

- ✓ **Maxwell SLS** model **is sufficient** whatever the method
- ✓ **GM2** → **over-parameterisation** of liver viscoelastic behaviour

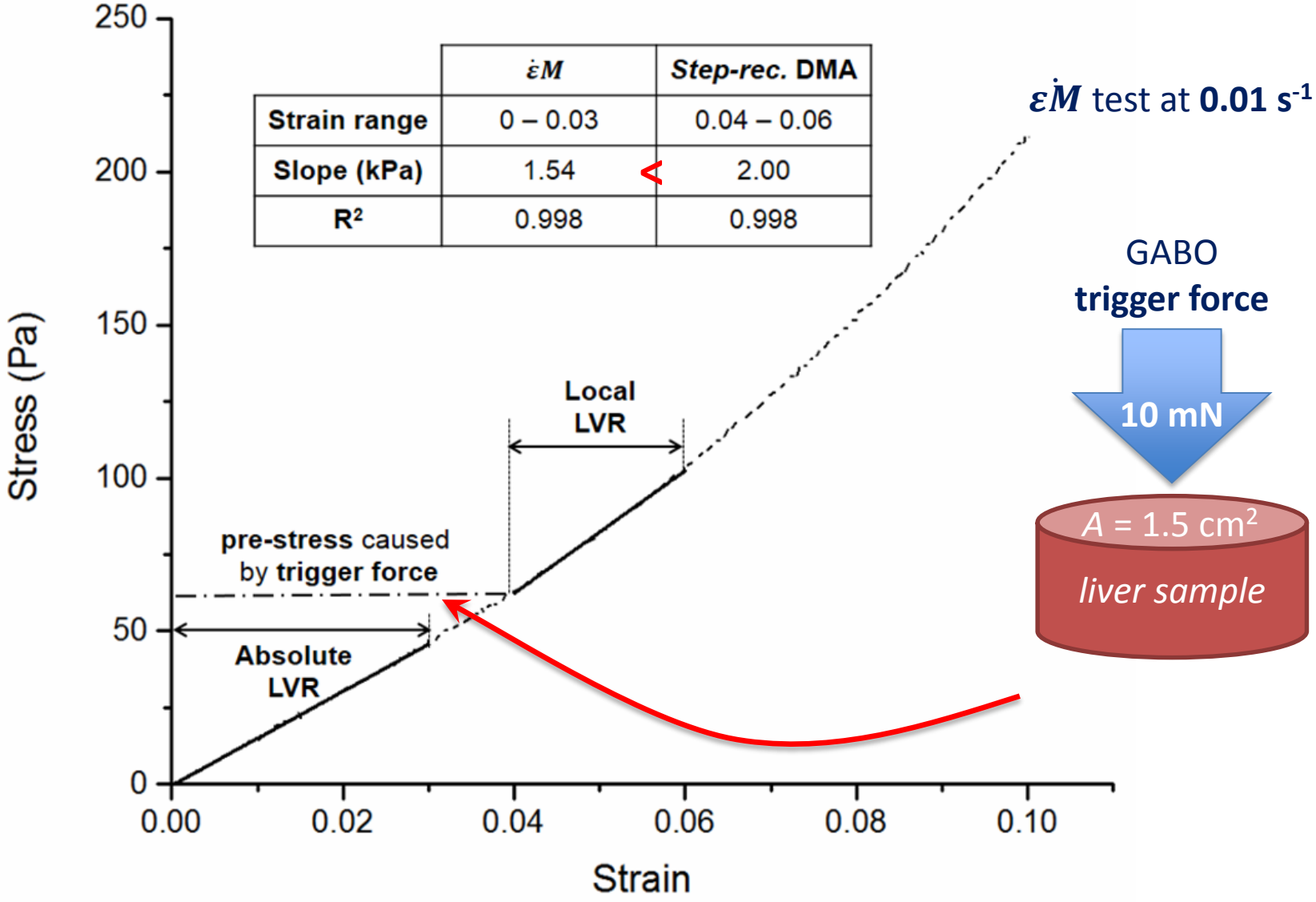


$\dot{\varepsilon}M$ and **SRDMA results** are **significantly different** (*t-test*, $p < 0.05$)

Absolute vs local LVR

	$\dot{\varepsilon}M$	Step-rec. DMA
Strain range	0 – 0.03	0.04 – 0.06
Slope (kPa)	1.54	2.00
R^2	0.998	0.998

$\dot{\varepsilon}M$ test at **0.01 s^{-1}**

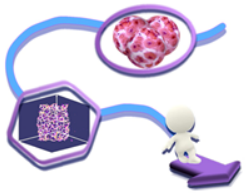


GABO
trigger force

10 mN

A = 1.5 cm^2

liver sample



Testing very soft tissues: conclusion

Long test
F or **strain trigger**

sample status changes
conventional DMA

Short test
F or **strain trigger**

local LVR
step-rec. DMA

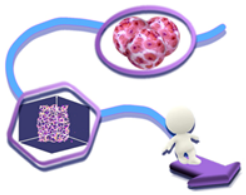
Short test
No trigger

actual properties
$\varepsilon\dot{M}$

- $\varepsilon\dot{M}$ gives a **good estimation** of **liver viscoelastic parameters in the LVR**
- A **wider range of** $\dot{\varepsilon}$ should be considered for a **more accurate estimation of** τ
- Caution in **over-interpreting *ex-vivo* data** (sample **status** is generally **different than *in-vivo*** and **dependent on many factors**, such as T, preservation period)

MY ACTIVITIES AT RESEARCH CENTRE «E. PIAGGIO»

ENGINEERING PATHOPHYSIOLOGICAL 3D IN-VITRO ORGAN MODELS

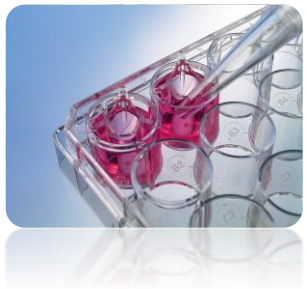


Project context: SoA

Animals

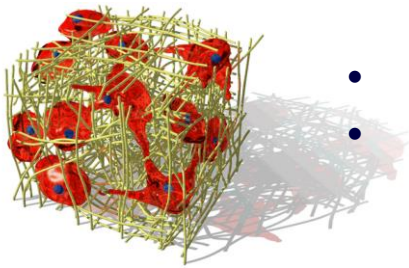
- **Scarcely representative** and **poorly predictive** for humans
- **Low benefit/cost** ratio and **not** suited for ***high-throughput*** testing
- **Ethically unacceptable**

In-vitro 2D



- **Standardised** platforms for **systematic, reliable** and **quantitative study of cell physiology, disease mechanism** and **drug efficacy**
- **Cells** on **2D substrates** behaves **differently than *in vivo***

In-vitro 3D

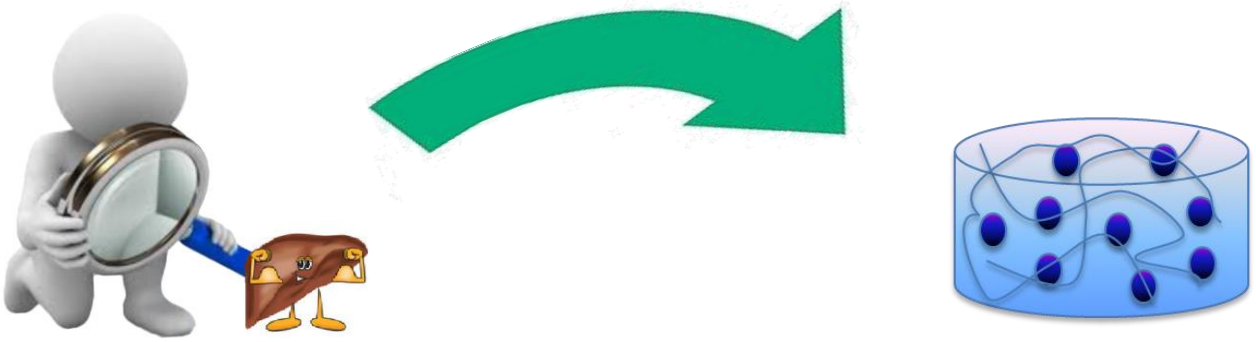


- **Better mimic** the **native cell micro-environment (ECM)**
- Provide **appropriate structural** and **functional support to cells** as well as **rational cues** for **diagnostic** and **therapeutic studies**

The strategy

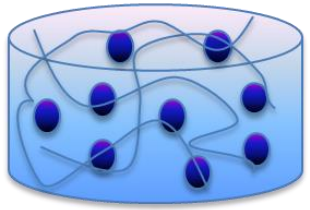
- **Tissue decellularisation** and **characterisation** to derive **ideal design specifications** for **ECM-mimicking scaffolds**
  - **Physicochemical characterisation**
  - **Histology**
  - **Mechano-structural analysis**

The strategy

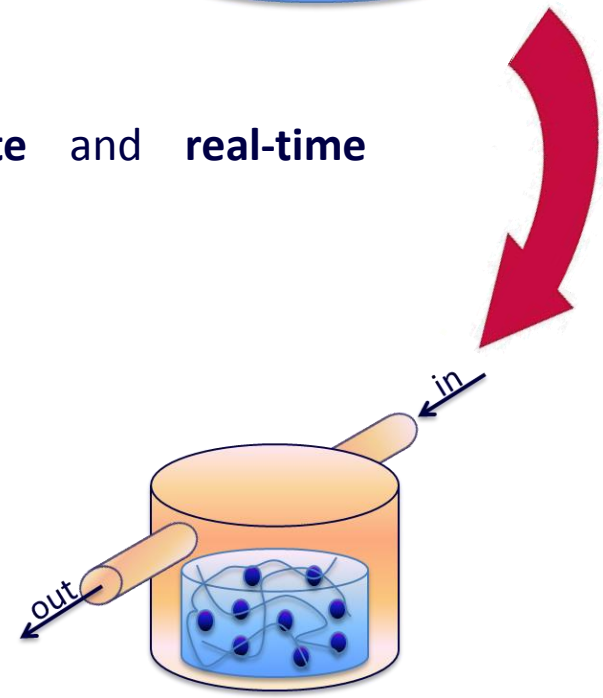


- **Modular approaches** to design **smart responsive biomimetic scaffolds** that:
  - **mimic key features** of the **native ECM**
  - **simulate tissue pathophysiology**
  - **provide** cultured cells with **specific cues** for the **selective study** of **cell behaviour and function**
  - **monitor** and **control 3D cellular environment**

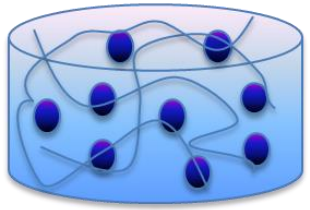
The strategy



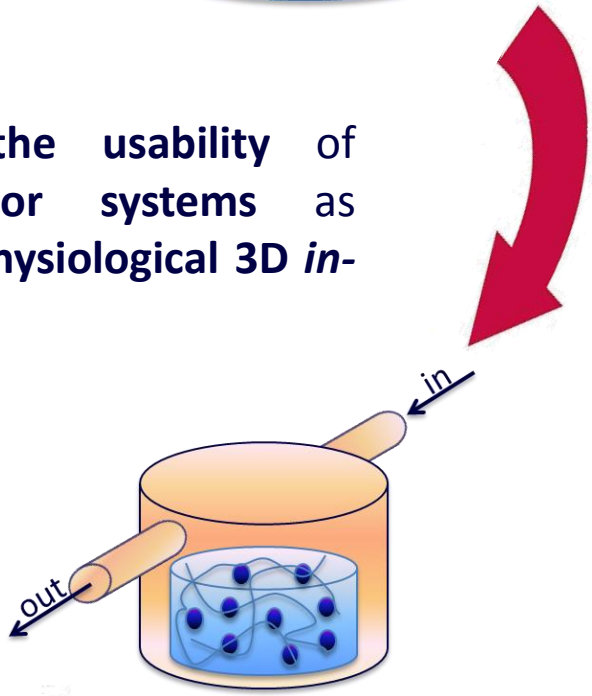
- Design **bioreactors** to **cultivate** and **real-time monitor** the **cellular constructs**

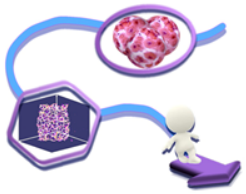


The strategy



- **Cell experiments** to **assess the usability** of developed **cell-scaffold-bioreactor systems** as platforms for **engineering pathophysiological 3D *in-vitro* organ models**

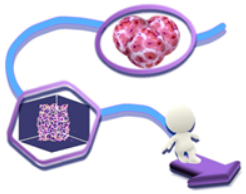




Bachelor thesis opportunities

- Liver **ECM-derived gels**
- **Brain clarification** and **slicing** for morphometric 3D analyses
- Acquisition and analysis of **human claustrum**
- Design of **low-cost devices** for **Africa project**
- Design and realisation of **mechanic** and **electronic components** for **new devices** for:
  - Cell cultures
  - Mechanical testing of tissues and biomaterials
  - Non-invasive real-time monitoring of 3D micro-environments
- **Calibration** of various **sensors** using **experimental** and **statistical methods**

http://www.centropiaggio.unipi.it/course/material/tesi-triennale.html



Contacts

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